

## Result

## 1 Project information

The basic information of the project is shown below:

Project ID: F23A430000062\_LEPjtwc0

• Product name: filter

• Sample size: 21

• Library type: DNBSEQ Eukaryotic mRNA sequencing - FFPE removed rRNA library

• Sequencing Platform: DNBseq

• Sequencing read Length: PE100

Clean fastq phred quality score encoding:Phred+33

## 2 Data production

After sequencing, the raw reads were filtered. Data filtering includes removing adaptor sequences, contamination and low-quality reads from raw reads.

Table 1 Statistics of clean data Sample Name Clean Reads Clean Base Read Length Q20(%) Q30(%) GC(%) C11 29,553,945 5,910,789,000 PE100 97.98 94.02 43.18 C12 28,876,958 5,775,391,600 97.74 93.53 43.12 PE100 C13 35,393,478 7,078,695,600 PE100 97.80 93.67 44.65 C14 28,414,142 5,682,828,400 PE100 97.80 93.69 44.49 C15 35,026,413 7,005,282,600 97.82 45.49 PE100 93.73 C16 35,085,674 7,017,134,800 PE100 97.85 93.81 44.76 C17 30,381,212 6,076,242,400 PE100 97.78 93.61 45.93 5,682,425,800 C18 28,412,129 PE100 97.92 94.03 45.56 C19 30,813,527 6,162,705,400 97.83 45.49 PE100 93.77 C2 32,253,105 6,450,621,000 PE100 98.02 94.08 43.51 C20 37,493,405 7,498,681,000 97.85 45.27 PE100 93.83 C21 40,972,452 8,194,490,400 PE100 98.00 94.30 51.93 C22 24,226,491 4,845,298,200 PE100 97.84 93.88 51.93 C23 37,324,542 7,464,908,400 PE100 97.87 93.86 45.58 **C**3 35,151,395 7,030,279,000 PE100 97.97 93.95 42.75 C4 40,389,920 8,077,984,000 PE100 98.14 94.47 42.81 34,158,579 6,831,715,800 98.06 42.91 **C**5 PE100 94.19 **C**6 32,564,989 6,512,997,800 PE100 98.12 94.41 42.76 **C7** 33,531,635 6,706,327,000 PE100 98.03 94.14 43.23 **C**8 25,417,116 94.64 43.51 5,083,423,200 PE100 98.20 **C**9 31,661,442 6,332,288,400 97.90 93.71 42.94 PE100

• Sample Name : Sample Name

Clean Reads: Clean reads
Clean Base: Clean bases
Read Length: Read length
Q20(%): Proportion of Q20

• Q30(%): Proportion of Q30

• GC(%): Proportion of GC

## 3 Quality control

The quality of data was examined after filtering.

# 3.1 The distribution of base percentage and qualities along reads after data filtering

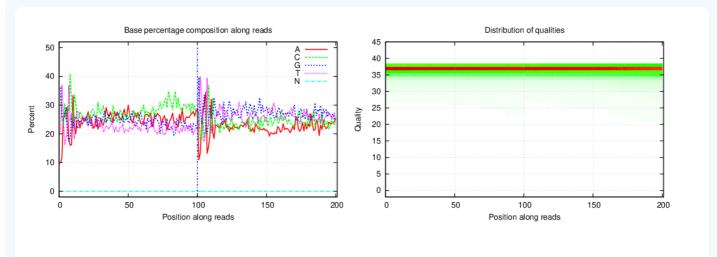


Figure 1 C21

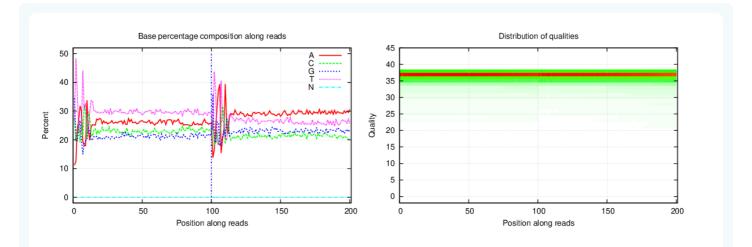


Figure 2 C13

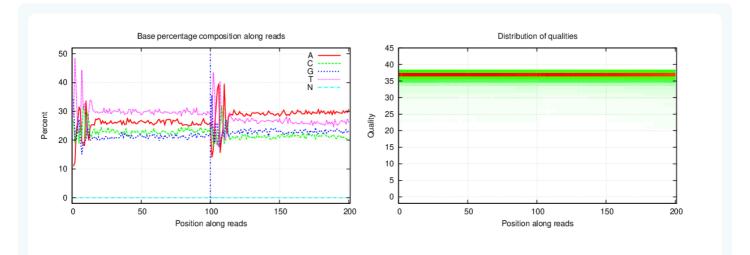


Figure 3 C14

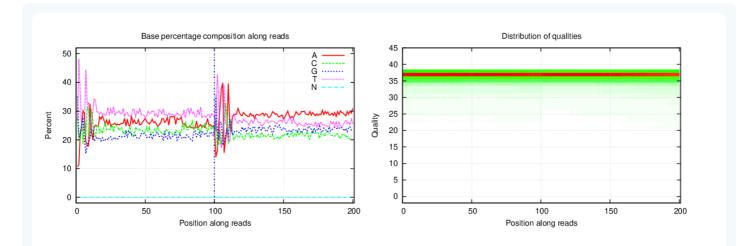


Figure 4 C20

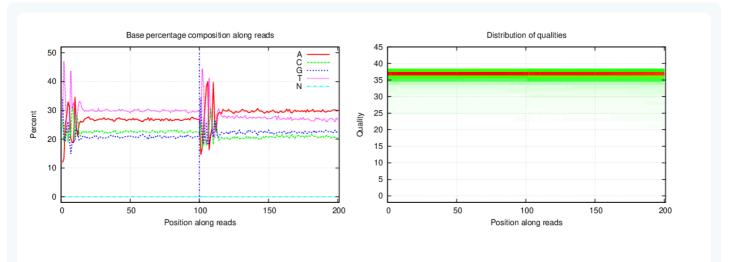


Figure 5 C2

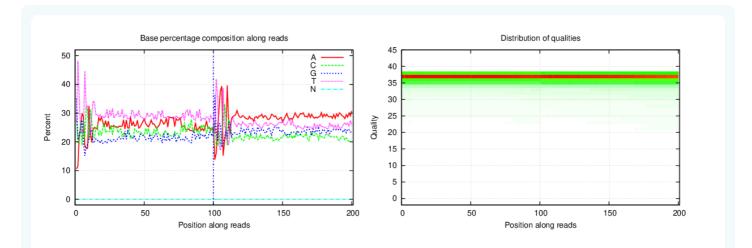


Figure 6 C19

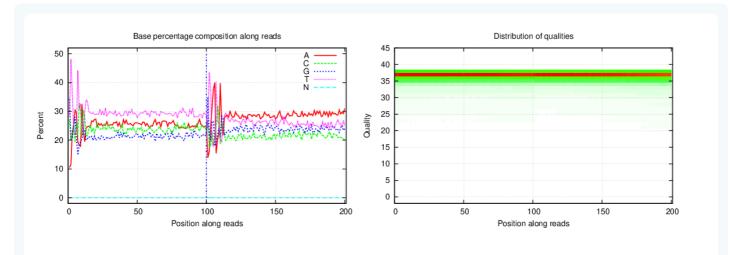


Figure 7 C15

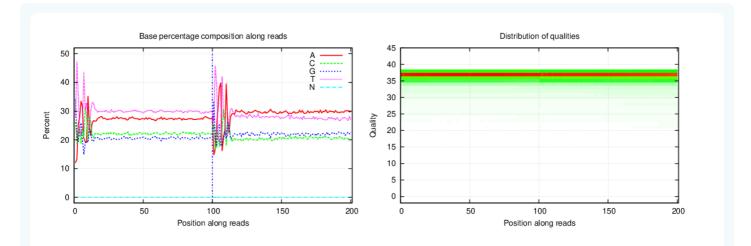


Figure 8 C9

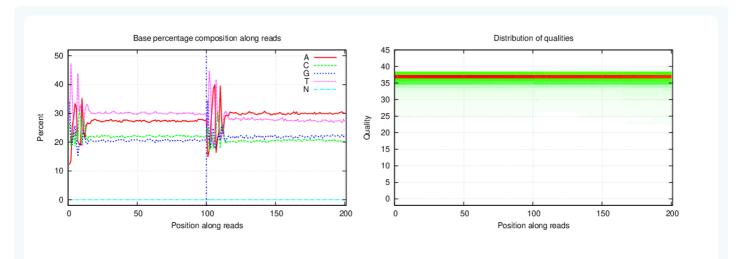


Figure 9 C4

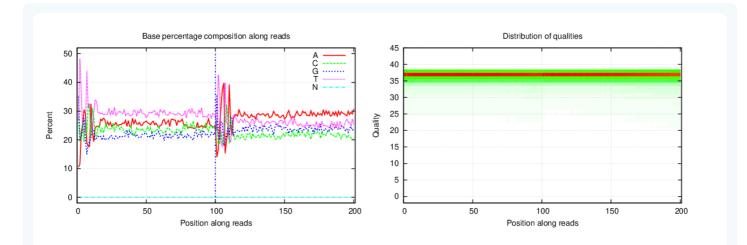


Figure 10 C18

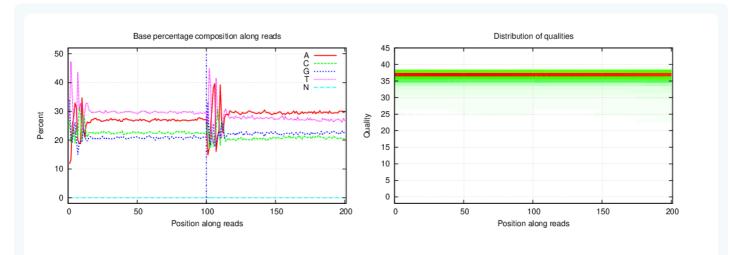


Figure 11 C8

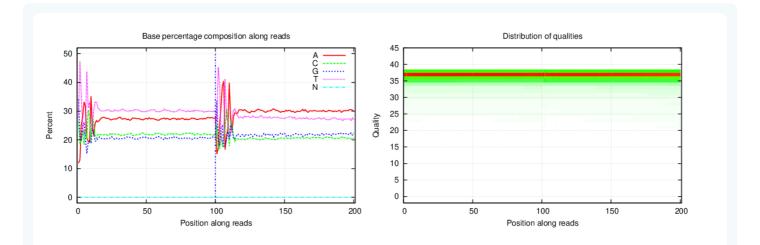


Figure 12 C3

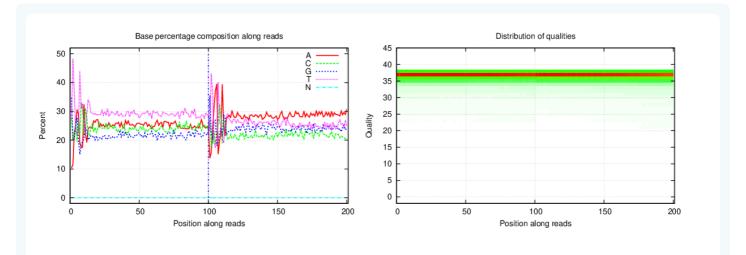


Figure 13 C17

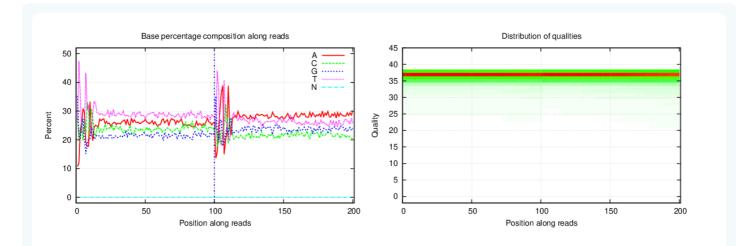


Figure 14 C23

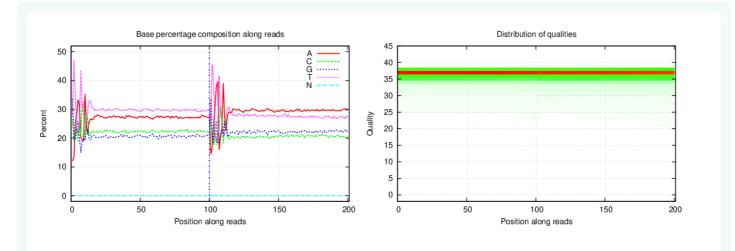


Figure 15 C11

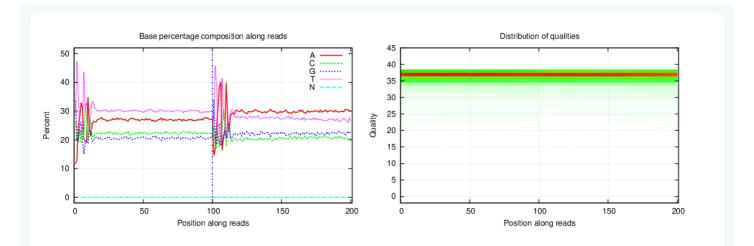


Figure 16 C12

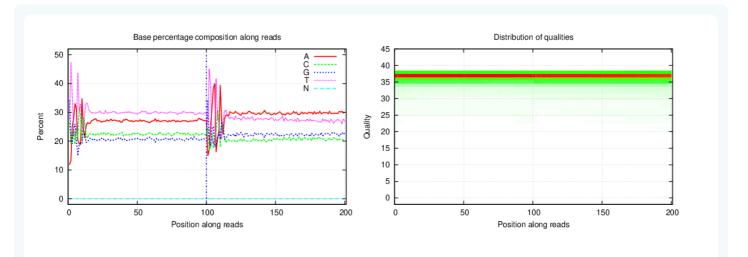


Figure 17 C7

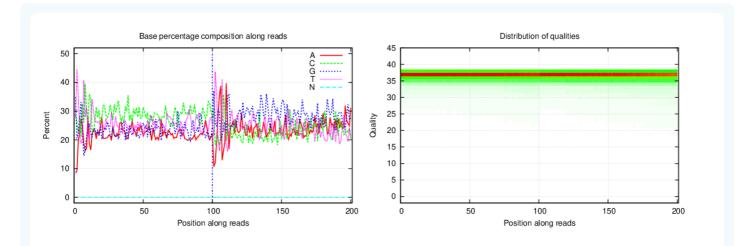


Figure 18 C22

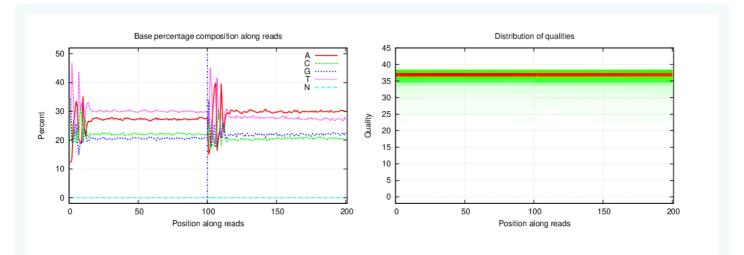


Figure 19 C5

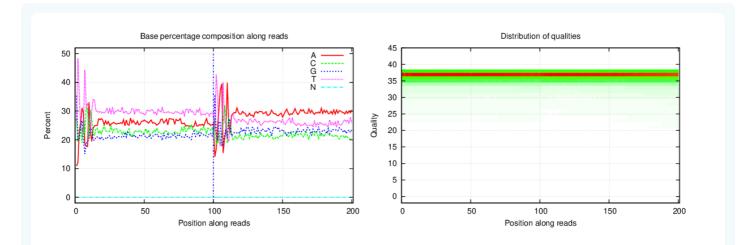


Figure 20 C16

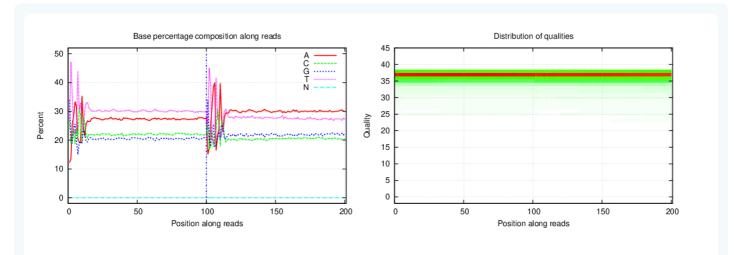


Figure 21 C6

## 4 References

[1] SOAPnuke: a MapReduce acceleration-supported software for integrated quality control and preprocessing of high-throughput sequencing data.

Chen Y, Chen Y, Shi C, et al.

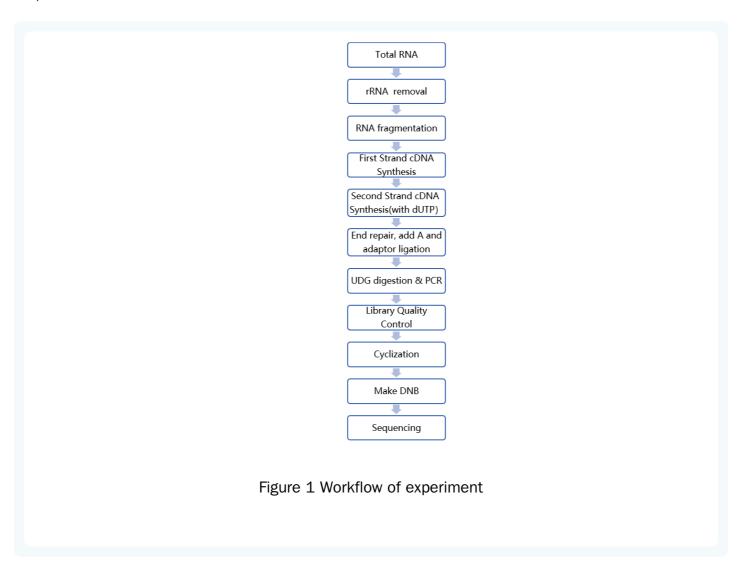
PMID: 29220494 PMCID: PMC5788068 DOI: 10.1093/gigascience/gix120



## Methods

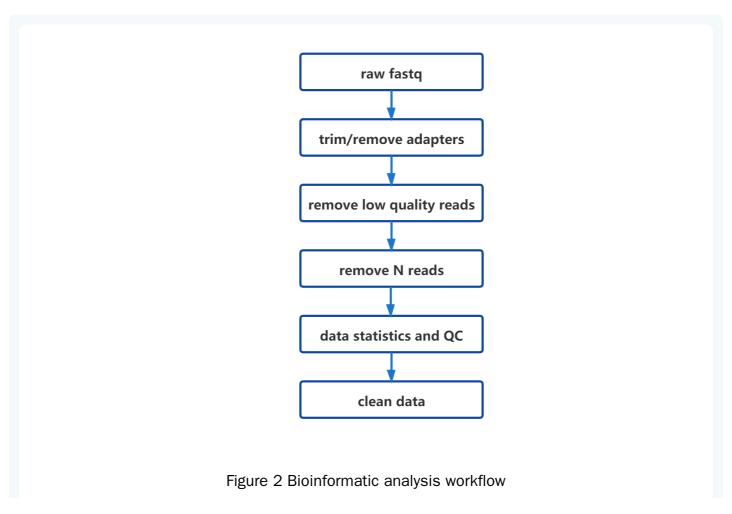
## 1 Experimental procedure

The library construction method and sequencing process are carried out according to the following steps:



- 1. Take a certain amount of total RNA samples, and use RNase H to remove rRNA;
- 2. After removal of rRNA, RNA molecules were fragmented into small pieces;
- 3. The fragmented RNA was synthesized into first strand cDNA using random primers;
- 4. The second strand cDNA was synthesized with dUTP instead of dTTP;
- 5. The synthesized cDNA was subjected to end-repair and 3' adenylated. Adaptors were ligated to the ends of these 3' adenylated cDNA fragments;
- 6. Digest the U-labeled second-strand template with Uracil-DNA-Glycosylase (UDG) and perform PCR amplification;
- 7. Library quality control;
- 8. Library circularization;
- 9. The library was amplified to make DNA nanoball (DNB);
- 10. Sequencing on DNBSEQ (DNBSEQ Technology) platform.

#### 2 Bioinformatic analysis workflow



## 3 Parameters for data filtering

Raw data with adapter sequences or low-quality sequences was filtered. We first went through a series of data processing to remove contamination and obtain valid data. This step was completed by SOAPnuke software developed by BGI.

SOAPnuke software filter parameters: " -n 0.001 -l 20 -q 0.4 --adaMR 0.25 --ada\_trim --minReadLen 100", steps of filtering:

- 1. Filter adapter: if the sequencing read matches 25.0% or more of the adapter sequence (maximum 2 base mismatches are allowed), cut the adapter;
- 2. Filter read length: if the length of the sequencing read is less than 100 bp, discard the entire read:
- 3. Remove N: if the N content in the sequencing read accounts for 0.1% or more of the entire read, discard the entire read:
- 4. Filter low-quality data: if the bases with a quality value of less than 20 in the sequencing read account for 40.0% or more of the entire read, discard the entire read;
- 5. Obtain Clean reads: the output read quality value system is set to Phred+33.

#### 4 References

[1] SOAPnuke: a MapReduce acceleration-supported software for integrated quality control and preprocessing of high-throughput sequencing data.

Chen Y, Chen Y, Shi C, et al.

PMID: 29220494 PMCID: PMC5788068 DOI: 10.1093/gigascience/gix120



# Help

#### 1 FASTQ format description

Images generated by sequencers are converted by base calling into nucleotide sequences, which are called raw data or raw reads and are stored in FASTQ format. FASTQ files are text files that store both reads sequences and their corresponding quality scores. Each read is described in four lines as follows:

@V350016857L4C001R0010000078/1

TTTTTCTGCTCCTTTTGATGCTATTAACAATTGCTTCAAGTTCAAGGGCACCTGCCTCAAAGTCCCTTTCTTCCAGACAAAATCTC

=, DDE@EFFF=DFDEFCCFDEFEGFEEAFDFFE=FFCFFEEEDFDEEEFDF8FFEFFEFF:FFEDF=EFDGE<1FDCEFFFFFDFE

The first line is the sequence identifier and related description information, starting with'@'; the second line is the base sequence information; the third line starts with'+', followed by the sequence identifier, description information, or nothing; The four lines are quality information, which corresponds to the sequence in the second line. Each base has a quality score. Depending on the scoring system, each character represents a different quality value.

The figure below shows the concise correspondence between the sequencing error rate and the sequencing quality value. Specifically, if the sequencing error rate is denoted by E and the base quality value is denoted by SQ, there are the following relationships:

$$SQ = -10 \times (log \frac{E}{1 - E})/(log 10)$$
 
$$E = \frac{Y}{1 + Y}$$
 
$$Y = \frac{SQ}{e^{-10 \times log 10}}$$

- 1. For the quality system data with a sequencing quality value of 33: the sequencing quality value of the base = the ASCII value corresponding to the quality information character -33, for example, the ASCII value corresponding to A is 65, then the corresponding base quality value is 65-33 = 32. The base quality value of the DNBSEQ sequencing platform ranges from 2 to 42.
- 2. For quality system data with a sequencing quality value of 64: the sequencing quality value of the base = the ASCII value corresponding to the quality information character -64, for example, the corresponding ASCII value of c is 99, then the corresponding base quality value is 99-64 =35. The base quality value of the DNBSEQ sequencing platform ranges from 2 to 43.

Table 1 A summary table of the relationship between sequencing error rate and sequencing
quality

5%       13       M       .         1%       20       T       5         0.1%       30       ^       ?	Sequencing error rate	Sequencing quality value	Character(Phred64)	Character(Phred33)
	5%	13	M	
0.1% 30 ^ ?	1%	20	Т	5
	0.1%	30	٨	?